

STRUCTURES OF OA-6129A, B₁, B₂ AND C, NEW CARBAPENEM ANTIBIOTICS

Takeo Yoshitaka, Ikuo Kojima, Kunio Isshiki, Azuma Watanabe, Yasutaka Shimauchi,
 Mitsuyasu Okabe, Yasuo Fukagawa and Tomoyuki Ishikura

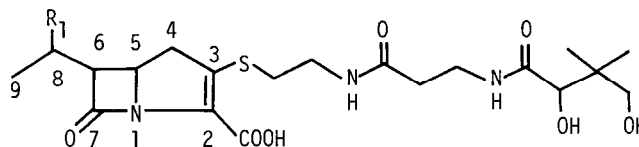
Central Research Laboratories, Sanraku-Ocean Co., Ltd., 9-1, Johnan 4-chome,
 Fujisawa 251, Japan

ABSTRACT The structures of carbapenems OA-6129A, B₁, B₂ and C were determined by spectroscopy and chemical transformation

In the course of our extensive work on carbapenems, new compounds OA-6129A(1), B₁(2), B₂(3) and C(4)¹⁾ have been isolated as sodium salts from *Streptomyces* sp. OA-6129. These compounds showed potent antimicrobial activity against Gram-positive and -negative bacteria²⁾ and inhibitory activity to β-lactamases. In this communication, we report the structure determination of the antibiotics by means of spectroscopic analysis, chemical degradation and direct derivation from carbapenem PS-5.

The u v and i r spectra of the four compounds strongly suggested that they had a carbapenem skeleton, a u v absorption maximum at 300 nm and i r absorption bands at 1750-1760(β-lactam carbonyl), 1660(amide carbonyl) and 1600 cm⁻¹(carboxylate anion) were very characteristic of the carbapenem structure^{3,4)}. The presence of a pantothenyl group in these

Fig 1



	R ₁	C-5	C-6	C-8	Pantoyl group
OA-6129A (1)	H	R	R	—	R
OA-6129B ₁ (2)	OH	R	R	*	R
OA-6129B ₂ (3)	OH	R	S	S	R
OA-6129C (4)	OSO ₃ H	R	R	*	R

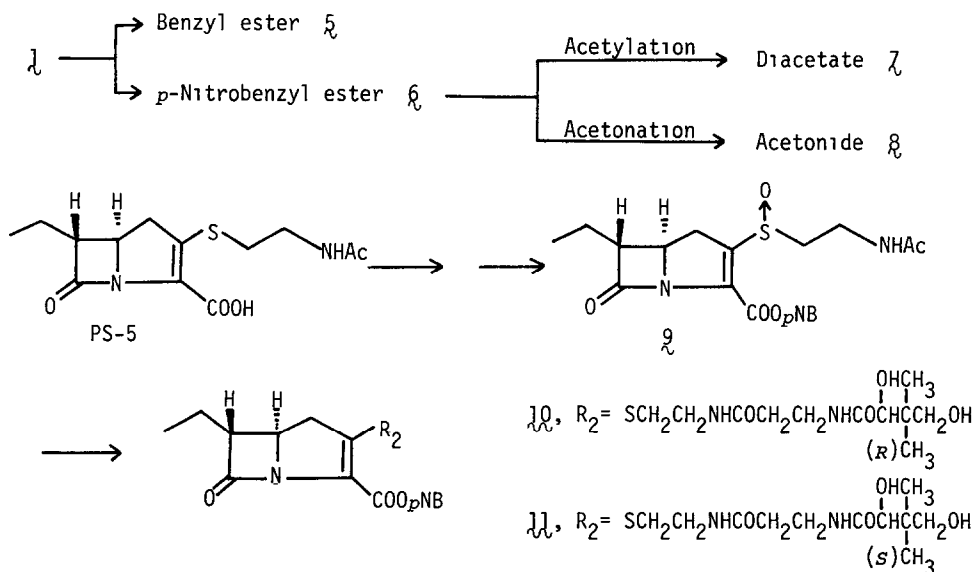
* not determined

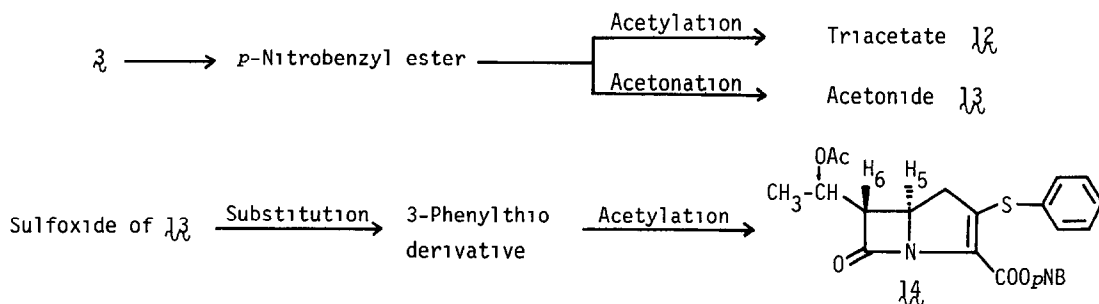
compounds was apparent from their n m r spectra⁵⁾ (two singlets attributed to geminal dimethyls at δ 0.86-0.92 and a triplet assigned to α -methylene attached to carbonyl at δ 2.45-2.48) and the formation of the corresponding acetonides and acetates. These findings clearly indicated that the four antibiotics had closely related structures including a carbapenem skeleton and a pantothenyl group.

The molecular formula of λ $C_{20}H_{31}N_3O_7S$ was deduced from the FD-MS spectrum [m/z 677 (M+1)] of the diacetate λ . Similarly to both cases of PS-5³⁾ and N-acetylthienamycin,⁴⁾ the characteristic u v absorption maximum of the sodium salt of λ at 300 nm shifted to 318-319 nm by esterification,^{6,7)} indicating that λ had the same chromophore as PS-5. Acid-catalyzed hydrolysis of λ (6N-HCl, 115°C) gave β -alanine. The mass spectrum of benzyl ester ξ ⁷⁾ revealed a strong peak at m/z 259 which showed the presence of cysteamine at C-3,⁸⁾ as found in PS-5 benzyl ester³⁾. Pantoate, β -alanine and cysteamine, taken together, clearly supported the presence of a pantetheinyl group at C-3⁸⁾ in λ . The side chain at C-6 was concluded to be ethyl by comparing the n m r signals of λ and PS-5. The *trans* configuration of the protons at C-5 and C-6 was confirmed by analysis of the lanthanide-induced n m r spectrum of the acetonide ρ , as the proton at C-5 appeared as a doublet of triplets ($J_{5,6}=3.0, J_{4,5}=9.0$ Hz). These data allowed us to conclude that λ was a derivative of PS-5 which had pantetheine at C-3⁸⁾ instead of N-acetylcysteamine.

The structure of λ was established by direct conversion of PS-5 to λ according to a method reported previously⁹⁾. S-Oxide of *p*-nitrobenzyl PS-5 ρ that was prepared from PS-5 by esterification and peracid oxidation was treated with (*r*)- and (*s*)-pantetheines to give λQ and λL , respectively. Product λQ was identical with ρ ⁶⁾ in every respect¹⁰⁾. Consequently, the structure of λ was unambiguously determined to be (5*R*,6*R*)-6-ethyl-7-oxo-3-[(*r*)-pantetheinyl]-1-azabicyclo[3.2.0]hept-2-ene-2-carboxylic acid.

The molecular formula of λ $C_{20}H_{31}N_3O_8S$ was also deduced from the FD-MS [m/z 735 (M+1)] of the triacetate λL . In the n m r spectrum⁵⁾ of λ 1-hydroxyethyl instead of C-6 ethyl in





λ was observed, while the remaining signals resembled very closely those of λ . The *trans* relationship between H-5 and H-6 was obvious from a coupling constant of $J_{5,6}=3.0$ Hz in the nmr spectrum of the acetate λ ¹¹⁾ which was derived from the acetonide λ by thiophenol replacement⁹⁾ via the S-oxide. The absolute configuration at C-8 bearing a hydroxy group was determined to be *s* by the Mosher's nmr-configuration correlation method¹²⁾ using the (*R*)-(+)- and (*S*)-(-)-MTPA esters^{4,13)} of λ . Thus, the structure of λ was (5,6-*trans*)-6-[(*S*)-1-hydroxyethyl]-7-oxo-3-pantetheinyl-1-azabicyclo[3.2.0]hept-2-ene-2-carboxylic acid.

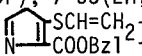
Compound λ , a minor component, was almost identical with λ in nmr⁵⁾, except for the signal of H-6 at δ 3.60. As the observed chemical shift and coupling constants were in good accordance with those reported for epithienamycin A,¹⁴⁾ λ was concluded to be an epimer of λ at C-6.

Elemental analysis gave a molecular formula of $C_{20}H_{31}N_3O_{11}S_2$ for λ ¹⁵⁾. The presence of a hydroxysulfonyloxy group in λ was indicated by an ir absorption band at $1220\text{--}1250\text{ cm}^{-1}$ and relatively high mobility on high voltage paper electrophoresis¹⁶⁾. Comparison of the nmr spectra⁵⁾ of λ and λ clearly showed that λ was a sulfate of λ , as both the methyl doublet and the H-8 multiplet shifted down-field in the spectrum of λ . The *cis* configuration between H-5 and H-6 was apparent from a coupling constant of $J_{5,6}=5.5$ Hz. Consequently, λ was decided to be (5,6-*cis*)-6-(1-hydroxysulfonyloxyethyl)-7-oxo-3-pantetheinyl-1-azabicyclo[3.2.0]hept-2-ene-2-carboxylic acid.

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5. NMR (D_2O , DSS) δ ppm: λ , 0.89 (3H, s), 0.92 (3H, s), 1.00 (3H, t, $J=7.5$ Hz), 1.60-2.00 (2H, m), 2.48 (2H, t, $J=6.5$ Hz), 2.80-3.65 (1H, m), 3.95 (2H, m). λ , 0.86 (3H, s), 0.89 (3H, s), 1.33 (3H, d, $J=6.0$ Hz), 2.47 (2H, t, $J=6.5$ Hz), 2.75-3.55 (10H, m), 3.60 (1H, dd, $J=5.0$ & 10.0 Hz), 3.93 (1H, s),

- 3 95-4 40(2H,m) δ , 0 87(3H,s), 0 92(3H,s), 1 28(3H,d, $J=7$ 0Hz), 2 45(2H,t, $J=6$ 5Hz),
 2 75-3 60(1H,m), 3 94(1H,s), 3 95-4 35(2H,m) δ , 0 86(3H,s), 0 89(3H,s), 1 49(3H,d,
 $J=6.5$ Hz), 2 47(2H,t, $J=7.0$ Hz), 2 70-3 60(10H,m), 3 83(1H,dd, $J=5$ 5&9 5Hz), 3 94(1H,s),
 4 10-4 43(1H,m), 4 78(1H,dq, $J=9$ 5&6 5Hz)
- 6 δ , UV λ_{\max} nm(ϵ) 319(8400), 270(10500), IR ν_{\max} cm^{-1} 1770(β -lactam), 1700(ester), 1665
 (amide), NMR(CD_2Cl_2 ,TMS) δ ppm 0.87(3H,s), 0.95(3H,s), 1 04(3H,t, $J=7$ 5Hz), 1 50-2 20
 (3H,m), 2 40(2H,t, $J=6$ 5Hz), 2 80-3 70(12H,m), 3 94(2H,m), 4 17(1H,br), 5 19(1H,d,
 $J=14$ 0Hz), 5 45(1H,d, $J=14$ 0Hz), 6 74(1H,br), 7 63(2H,d, $J=9$ 0Hz), 8 18(2H,d, $J=9$ 0Hz)
- 7 δ , UV λ_{\max} nm(ϵ) 318(7400), MS(m/z) 259[⁺, 242
- 8 In this paper we employed the numbering system as shown in Fig 1
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10. $[\alpha]_D^{24}$ (c 1 0, CH_2Cl_2), δ +52 4°, $[\eta]$ +51 7°, $[\mu]$ +30 3°
- 11 δ , NMR(CDCl_3 ,TMS) δ ppm 1 36(3H,d, $J=6$ 5Hz), 2 03(3H,s), 2 66(2H,d, $J=9$ 0Hz), 3 28
 (1H,dd, $J=3$ 0&4 5Hz), 3 92(1H,dt, $J=3$ 0&9 0Hz), 4 95-5 40(2H,m), 5 50(1H,d, $J=14$ 0Hz),
 7 20-7 57(5H,m), 7 60(2H,d, $J=9$ 0Hz), 8 15(2H,d, $J=9$ 0Hz)
- 12 J A.Dale & H S Mosher, J Amer Chem Soc 95, 512(1973)
- 13 NMR(CDCl_3 ,TMS) δ ppm, (R)-(+) -MTPA ester 0 94(3H,s), 1 00(3H s), 1 39(3H,s), 1 43(3H,s),
 1 52(3H,d, $J=7$ 5Hz,H-9), 2 43(2H,t, $J=6$ 5Hz), 2 80-3 75(14H,m), 3 96(1H,m), 4 03(1H,s),
 5 22(1H,d, $J=14$ 0Hz), 5 40(1H,m), 5 41(1H,d, $J=14$ 0Hz), 6 48(1H,br), 6 87(1H,br), 7 25-
 7 64(7H,m), 8 12(2H,d, $J=8$ 0Hz) (S)-(-) -MTPA ester 0 93(3H,s), 1 00(3H,s), 1 39(3H,s),
 1 42(3H,s), 1 42(3H,d, $J=7$ 5Hz,H-9), 2 43(2H,t, $J=6$ 5Hz), 2 68-3 85(14H,m), 4 03(1H,s),
 4 05(1H,m), 5 22(1H,d, $J=14$ 0Hz), 5 43(1H,d, $J=14$ 0Hz), 5 45(1H,m), 6 47(1H,br), 6 90
 (1H,br), 7 22-7 70(7H,m), 8 12(2H,d, $J=8$ 0Hz)
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- 15 Elemental analysis ($\text{C}_{20}\text{H}_{29}\text{N}_3\text{O}_{11}\text{S}_2\text{Na}_2 \cdot \text{H}_2\text{O}$), Found C 39 18%, H 4 91%, N 6 86%, S 10 08%
 Calcd C 39 02%, H 5 08%, N 6 83%, S 10 42%
- 16 Rm PS-5, 1 69 for δ , 0 67 for δ , δ & δ at pH 8 4

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